

# Package ‘MSiP’

July 21, 2025

**Type** Package

**Title** 'MassSpectrometry' Interaction Prediction

**Version** 1.3.7

**Description** The 'MSiP' is a computational approach to predict protein-protein interactions from large-scale affinity purification mass 'spectrometry' (AP-MS) data. This approach includes both spoke and matrix models for interpreting AP-MS data in a network context. The ``spoke" model considers only bait-prey interactions, whereas the ``matrix" model assumes that each of the identified proteins (baits and prey) in a given AP-MS experiment interacts with each of the others. The spoke model has a high false-negative rate, whereas the matrix model has a high false-positive rate. Although, both statistical models have merits, a combination of both models has shown to increase the performance of machine learning classifiers in terms of their capabilities in discrimination between true and false positive interactions.

**Depends** R (>= 3.6.0)

**Imports** dplyr (>= 1.0.6), tibble (>= 3.1.2), tidyr (>= 1.1.3),  
magrittr (>= 2.0.1), plyr (>= 1.8.6), PRROC (>= 1.3.1), caret  
(>= 6.0.88), e1071 (>= 1.7.7), mice (>= 3.13.0), pROC (>=  
1.17.0.1), ranger (>= 0.12.1)

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.1.1

**Suggests** knitr, markdown

**VignetteBuilder** knitr

**NeedsCompilation** no

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**Repository** CRAN

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cPASS

*cPASS*

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### Description

This function applies Comparative Proteomic Analysis Software Suite (CompPASS) model to score instances (e.g., bait-prey interactions (BPIs) in the data.frame. The CompPASS is a robust statistical scoring scheme for assigning confidence scores to bait-prey interactions (Sowa et al., 2009). This function was based on the source code. <https://github.com/dnusinow/cRomppass>

### Usage

```
cPASS(datInput)
```

### Arguments

`datInput` Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).

### Value

Data frame containing bait-prey pairs with average peptide spectrum match (PSMs), total PSMs, ratio total PSMs, Z-score, S-score, D-score and WD-score.

### Author(s)

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

### References

Huttlin, E. L., Ting, L., Bruckner, R. J., Gebreab, F., Gygi, M. P., Szpyt, J., et al. (2015). The BioPlex Network: A Systematic Exploration of the Human Interactome. *Cell* 162, 425-440.

Sowa, M. E., Bennett, E. J., Gygi, S. P., and Harper, J. W. (2009). Defining the human deubiquitinating enzyme interaction landscape. *Cell* 138, 389-403.

**Examples**

```
data(SampleDatInput)
datScoring <- cPASS(SampleDatInput)
head(datScoring)
```

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diceCoefficient	<i>diceCoefficient</i>
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**Description**

This function applies Dice coefficient to score instances (e.g., bait-prey interactions (BPIs) in the data.frame. The Dice coefficient was first applied by Zhang et al., 2008 to score interactions between all identified proteins (baits and preys) in a given AP-MS experiment.

**Usage**

```
diceCoefficient(datInput)
```

**Arguments**

datInput      Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).

**Value**

Data frame containing bait-prey pairs with Dice coefficient score, a number between 0 and 1

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**References**

Zhang, B., Park, B.-H., Karpinets, T., and Samatova, N. F. (2008). From pull-down data to protein interaction networks and complexes with biological relevance. *Bioinformatics* 24, 979-986.

**Examples**

```
data(SampleDatInput)
datScoring <- diceCoefficient(SampleDatInput)
head(datScoring)
```

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jaccardCoefficient	<i>jaccardCoefficient</i>
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**Description**

This function computes the Jaccard similarity coefficient scores for instances (e.g., bait-prey interactions (BPIs)) in the data.frame.

**Usage**

```
jaccardCoefficient(datInput)
```

**Arguments**

datInput	Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).
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**Value**

Data frame containing bait-prey pairs with the Jaccard coefficient score, a number between 0 and 1

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**Examples**

```
data(SampleDatInput)
datScoring <- jaccardCoefficient(SampleDatInput)
head(datScoring)
```

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overlapScore	<i>overlapScore</i>
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**Description**

This function computes the overlap similarity scores for instances (e.g., bait-prey interactions (BPIs)) in the data.frame.

**Usage**

```
overlapScore(datInput)
```

**Arguments**

datInput	Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).
----------	---

**Value**

Data frame containing bait-prey pairs with the overlap score, a number between 0 and 1

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**References**

Nepusz, T., Yu, H., and Paccanaro, A. (2012a). Detecting overlapping protein complexes in protein-protein interaction networks. *Nat. Methods* 9, 471.

**Examples**

```
data(SampleDatInput)
datScoring <- overlapScore(SampleDatInput)
head(datScoring)
```

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rfTrain

*rfTrain*

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**Description**

The labeled feature matrix can be used as input for Random Forest (RF) classifier. The classifier then assigns each bait-prey pair a confidence score, indicating the level of support for that pair of proteins to interact. Hyperparameter optimization can also be performed to select a set of parameters that maximizes the model's performance. This function also computes the areas under the precision-recall (PR) and ROC curve to evaluate the performance of the classifier.

**Usage**

```
rfTrain(
  dtInput,
  impute = TRUE,
  p = 0.3,
  parameterTuning = TRUE,
  mtry = seq(from = 1, to = 10, by = 2),
  min_node_size = seq(from = 1, to = 9, by = 2),
  splitrule = c("gini"),
  metric = "Accuracy",
  resampling.method = "repeatedcv",
  iter = 5,
  repeats = 5,
  pr.plot = TRUE,
  roc.plot = TRUE
)
```

**Arguments**

<code>dtInput</code>	Data frame containing instances with class labels
<code>impute</code>	Logical value, indicating whether to impute missing values
<code>p</code>	The percentage of data that goes to training; defaults to 0.3
<code>parameterTuning</code>	Logical value; indicating whether to tune rf hyper parameters
<code>mtry</code>	Number of variables to possibly split at in each node and it is bound by the number of variables in your model
<code>min_node_size</code>	Minimal node size
<code>splitrule</code>	Splitrule rule for classification: 'gini', 'extratrees' or 'hellinger' with default 'gini'
<code>metric</code>	A string that specifies what summary metric will be used to select the optimal model; default to Accuracy
<code>resampling.method</code>	The resampling method: 'boot', 'boot632', 'optimism_boot', 'boot_all', 'cv', 'repeatedcv', 'LOOCV', 'LGOVCV'; defaults to repeatedcv
<code>iter</code>	Number of resampling iterations; defaults to 5
<code>repeats</code>	for repeated k-fold cross validation only; defaults to 5
<code>pr.plot</code>	Logical value, indicating whether to plot precision-recall (PR) curve
<code>roc.plot</code>	Logical value, indicating whether to plot ROC curve

**Value**

Data frame containing a classification results for all instances in the data set, where positive confidence score corresponds to the level of support for the pair of proteins to be true positive, whereas negative score corresponds to the level of support for the pair of proteins to be true negative.

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**Examples**

```
data(testdfClassifier)

predidcted_RF <-
rfTrain(testdfClassifier,impute = FALSE, p = 0.3, parameterTuning = FALSE,
mtry = seq(from = 1, to = 5, by = 1),
min_node_size = seq(from = 1, to = 5, by = 1),
splitrule =c("gini"),metric = "Accuracy",
resampling.method = "cv",iter = 2,repeats = 2,
pr.plot = TRUE, roc.plot = FALSE)
head(predidcted_RF)
```

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SampleDatInput	<i>Test data for scoring</i>
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**Description**

Bait-Prey Interactions (BPIs)

**Usage**

```
data(SampleDatInput)
```

**Details**

- Experiment ID
- Replicate
- Bait
- Prey
- counts

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

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simpsonCoefficient	<i>simpsonCoefficient</i>
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**Description**

This function computes the Simpson similarity coefficient scores for instances (e.g., bait-prey interactions (BPIs)) in the data.frame.

**Usage**

```
simpsonCoefficient(datInput)
```

**Arguments**

datInput	Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).
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**Value**

Data frame containing bait-prey pairs with Simpson coefficient, a number between 0 and 1

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**Examples**

```
data(SampleDatInput)
datScoring <- overlapScore(SampleDatInput)
head(datScoring)
```

---

 svmTrain

*svmTrain*


---

**Description**

The labeled feature matrix can be used as input for Support Vector Machines (SVM) classifier. The classifier then assigns each bait-prey pair a confidence score, indicating the level of support for that pair of proteins to interact. Hyperparameter optimization can also be performed to select a set of parameters that maximizes the model's performance. This function also computes the areas under the precision-recall (PR) and ROC curve to evaluate the performance of the classifier.

**Usage**

```
svmTrain(
  dtInput,
  impute = TRUE,
  p = 0.3,
  parameterTuning = TRUE,
  cost = seq(from = 2, to = 10, by = 2),
  gamma = seq(from = 0.01, to = 0.1, by = 0.02),
  kernel = "radial",
  ncross = 10,
  pr.plot = TRUE,
  roc.plot = TRUE
)
```

**Arguments**

dtInput	Data frame containing instances with class labels
impute	Logical value, indicating whether to impute missing values
p	The percentage of data that goes to training; defaults to 0.3
parameterTuning	Logical value; indicating whether to tune SVM hyper parameters
cost	Cost of constraints violation
gamma	Parameter needed for all kernels except linear
kernel	Kernel type: 'linear', 'polynomial', 'sigmoid', or 'radial'; defaults to 'radial'



ncross	K-fold cross validation on the training data is performed to assess the quality of the model; defaults to 10
pr.plot	Logical value, indicating whether to plot precision-recall (PR) curve
roc.plot	Logical value, indicating whether to plot ROC curve

**Value**

Data frame containing a classification results for all instances in the data set, where positive confidence score corresponds to the level of support for the pair of proteins to be true positive, whereas negative score corresponds to the level of support for the pair of proteins to be true negative.

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

**Examples**

```
data(testdfClassifier)
predidcted_SVM <-
svmTrain(testdfClassifier,impute = FALSE,p = 0.3,parameterTuning = FALSE,
cost = seq(from = 2, to = 10, by = 2),
gamma = seq(from = 0.01, to = 0.10, by = 0.02),
kernel = "radial",ncross = 10,
pr.plot = FALSE, roc.plot = TRUE)
head(predidcted_SVM)
```

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testdfClassifier	<i>Test data for classifier</i>
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**Description**

Scored data.frame

**Usage**

```
data(testdfClassifier)
```

**Author(s)**

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

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Weighted.matrixModel *Weighted.matrixModel*

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### Description

This function computes the weighted matrix model for instances (e.g., bait-prey interactions (BPIs)) in the data.frame. The output of the weighted matrix model includes the number of experiments for which the pair of proteins is co-purified (i.e., k) and  $-1 \cdot \log(\text{P-value})$  of the hypergeometric test (i.e., logHG) given the experimental overlap value, each protein's total number of observed experiments, and the total number of experiments (Drew et al., 2017).

### Usage

```
Weighted.matrixModel(datInput)
```

### Arguments

datInput            Data frame with column names: Experiment.id, Replicate, Bait, Prey, and count (i.e., prey count).

### Value

Data frame containing bait-prey pairs with k (i.e., number of co-purifications) & logHG (i.e.,  $-1 \cdot \log(\text{P-val})$  of the hypergeometric test)

### Author(s)

Matineh Rahmatbakhsh, <matinerb.94@gmail.com>

### References

Drew, K., Lee, C., Huizar, R. L., Tu, F., Borgeson, B., McWhite, C. D., et al. (2017). Integration of over 9,000 mass spectrometry experiments builds a global map of human protein complexes. *Mol. Syst. Biol.* 13, 932.

### Examples

```
data(SampleDatInput)
datScoring <- Weighted.matrixModel(SampleDatInput)
head(datScoring)
```

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